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Microbiological Characteristics of Vacuum-CO, - Packaged Broiler Carcasses

Bulk packaging of fresh broiler products (whole or part) in ice permits distribution from the South to major markets in urpanized areas throughout the United States, but shelf life of icepacked poultry is short. Therefore, an alternative to ice packing is needed to (1) extend shelf life, (2) reduce transportation costs by replacing ice with salable products and (3) facilitate physical handling by eliminating melting ice.

Vacuum packing has been inroduced as an alternative. Canoni and Bolther (1973) identified several advantages of vacuum packaging, including reduction in weight loss due to dehydration, preservation of muscle color in its freshest state and elimination of external contamination. In addi-

Procedure

Sample Preparation and Storage

Ice packed and vacuum- CO₂packed broiler carcasses (with giblet packages) were obtained from a commercial processing plant. The ice-packed samples were prepared by placing broiler carcasses directly into crushed ice. The vacuum- CO_2 - packed samples were prepared by placing three broiler carcasses in 4.0 mil polyethylene pouches and sealing after drawing vacuum (44.4 cm or 17.5 in) and adding CO₂ at rates of 3.54 x 10⁻⁴ to $3.97 \text{x} 10^{-4} \text{m}^{3}/\text{kg}$ carcass. Ten samples of each treatment were stored in -2.2 to 1.1°C and 1.1 to 3.3°C refrigerators in the Mississippi State University Poultry Science Laboratory. One sample from each storage condition was selected randomly for analysis at each testing period. Experiments were repeated.

tion, vacuum packaging improves organoleptic acceptability of ground beef (Jaye *et al.*, 1962).

Arafa and Chen (1975) observed that vacuum packaging of cut-up fresh broilers did not prolong shelf life during refrigerated storage. Microorganisms responsible for the spoilage of vacuum-packed, cutup fresh broilers were members of *Enterobacter* species, and large numbers of *Pseudomonas* species were found in control samples.

Treatment with carbon dioxide (CO_2) is another way to improve packaging. CO_2 in the environment selectively inhibits growth of bacteria. Researchers have found that a combination of low temperature (0-10°C) and CO_2 in a controlled environment effectively extends the shelf life of red meat

(Baran *et al.*, 1970), poultry (Wabeck *et al.*, 1968) and pork (Adams and Huffman, 1972). Recently, Sander and Soo (1978) reported that substituting 95% CO_2 for the air in packages of fresh broilers limited the growth of aerobic and anaerobic microorganisms. They indicated that packaging broiler carcasses in Nylon/Surlyn film with carbon dioxide addition rates of 3.6×10^{-4} and $7.22 \times 10^{-4} \text{m}^3/\text{kg}$ carcass, respectively, extended shelf life to 22 and 27 days at 1.1°C .

The purpose of this study was to evaluate the effect of vacuum- CO_2 -packaging on the shelf life of broilers under two different storage temperatures.

Drip Measurement

The vacuum-CO₂ packages were cut at the corner and drained for 30 seconds. The drip was weighed and converted to a percentage of original carcass weight.

Sampling of Microorganisms

Carcass - - - Samples for microbiological study were obtained by swabbing the breast skin with a moist, sterile cotton swab (Falcon plastics, CA) for thirty seconds in a different direction. The skin area swabbed was 1 sq inch (6.45 cm²) circumscribed by a sterile aluminum foil template. The cotton tip was broken into a serial dilution containing 10 ml of nutrient broth, and the solution was shaken vigorously to disperse the cotton swab and thoroughly mix the microflora.

Gizzard---The gizzard sample

was weighed into a sterile, widemouth 250 ml Erlenmeyer flask. An equal weight of sterile nutrient broth was added, and the mixture was shaken vigorously for one minute.

Plate Counts:

Plate counts were made in duplicate by using standard method agar (BBL). Plates were incubated for 72 hours at 20°C to determine the psychrophilic counts. For anaerobic counts, anaerobic agar (BBL) was used, and plates were incubated for 48 hours at 37°C. Average counts from the duplicate plates were reported as the number of bacteria per cm² of skin surface or number of bacteria per gram of gizzard.

Coliform Most Probable Number (MPN)

Methods for the coliform test as

described by Thatcher and Clark (1968) were followed.

Psychrotrophic count plates of the zero time and spoiled carcasses were used for selecting isolates under the two packing conditions. Carcasses stored for 28 days at 1.1 to 3.3°C were used as spoiled samples. All colonies from plates

RESULTS AND DISCUSSION

The log of total bacterial counts on 6.5 cm² of skin surface at 20°C, suggested by Elliott and Michener as a spoilage indicator, revealed that vacuum-CO ₂ - packed broiler carcasses spoiled after 21 days of storage at 1.1 to 3.3°C. Ice-packed samples stored under the same conditions spoiled after 14 days of storage. Lowering the storage temperature to -2.2 to 1.1°C extended the shelf life for the vacuumcontaining 10 to 30 colonies were isolated without bias and were purified by a streak-plate technique.

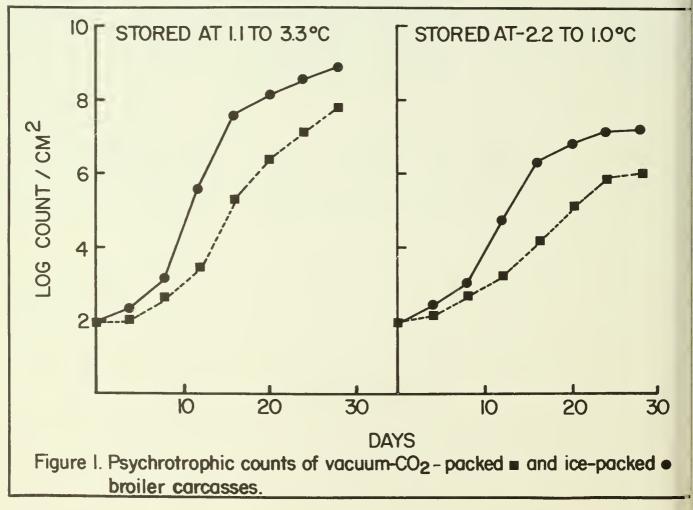
Identification of Microorganisms:

The purified cultures were transferred to standard method agar slants and incubated at 20°C, and these fresh cultures were used for identification. Gram sta motility, oxidative and ferme tative metabolism, cytochron oxidase and catalase tests of the cultures were made according the methods described by Colli and Lyne (1970). The scheme Freeman *et al* (1976) was used identify the isolates.

CO₂ - packed and the ice-packed broiler carcasses about five and four days, respectively.

The finding of longer shelf life for broiler carcasses in a CO_2 atmosphere agreed with the results of Killeffer (1930) and Coyne (1932). They determined that meat and fish products can be kept fresh longer when refrigerated in a CO₂ atmosphere. Our results supported the findings of Sander and Soo (1978) who indicate that, at storage temperature of 1.1° packaging broiler carcasses Nylon/Surlyn film with $3.6 \times 10^{\circ}$ and $7.22 \times 10^{-4} \text{m}^3$ CO $_2$ /kg carca extended the shelf life eight to to days, respectively.

The growth of psychrotrophilmicroorganisms (Figure 1) with delayed by the absence of a and/or the presence of 3.54×10^{-4} ; $3.97 \times 10^{-4} \text{m}^3 \text{CO}_2$ /kg carcass. Ps,



nrotrophic counts were constently lower for the vacuum- O_2 -packed samples than for the e-packed controls. A longer rowth lag phase was observed for ie vacuum-CO₂ - packed samples nan for the ice-packed control. imilar trends were observed at oth storage temperature ranges. otal aerobic counts for both ackaging methods were lower for amples stored at -2.2 to 1.1°C than or those stored at 1.1 to 3.3°C. imilar trends were reported by Bailey et al (1979) for broilers tored at 2°C and 5°C.

The initial psychrophilic counts or both types of broiler carcasses vere lower than those of the psyhrotrophic counts (Figures 1 and 2); however, the psychrophilic counts were slightly higher than the psychrotrophic counts after 16 days of storage.

Anaerobic counts increased steadily for both treatments. Lower storage temperature resulted in lower anaerobic counts for the carcasses. Lower anaerobic counts were observed for the vacuum- CO_2 – packed than for the ice-packed controls (Figure 3).

A log microbial count of 7.49/gram suggested by Charoenpong and Chen (1979) as detectable spoilage level for refrigerated gizzards, indicated that gizzards packed in giblet bags and stuffed in broiler cavities spoiled after 17 days of storage at 1.1 to 3.3°C. Log total counts/gram of gizzard were higher than log counts/cm² of the broiler skin. Thus, the vacuum- CO_2 -packaging condition affected the growth of microorganisms on gizzards more than it did those of the carcass (Figures 1 and 4).

Under the same storage conditions, vacuum-CO₂ packaging extended the shelf life of gizzards in giblet bags about eight days beyond that of ice pack. Gizzard samples from the vacuum-CO₂packaged carcasses stored at -2.2 to 1.1° C had counts of less than $1.4x10^{5}$ cells/gofgizzard at 28 days of storage (Figure 4). No off-odor was detected for these gizzard samples.

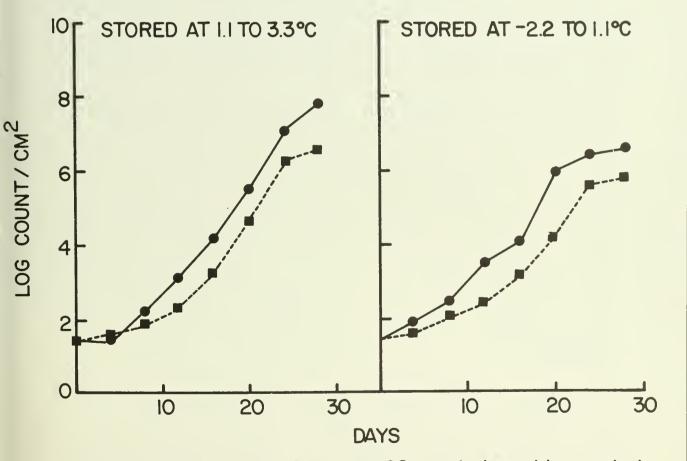
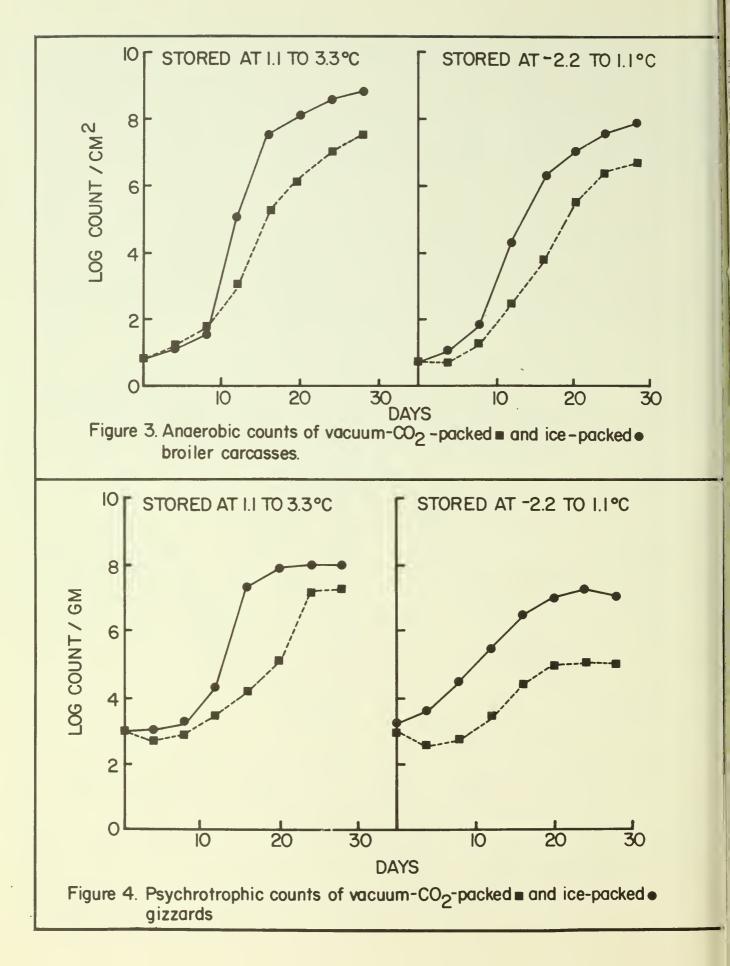


Figure 2. Psychrophilic counts of vacuum - CO₂-packed and ice-packed broiler carcasses.



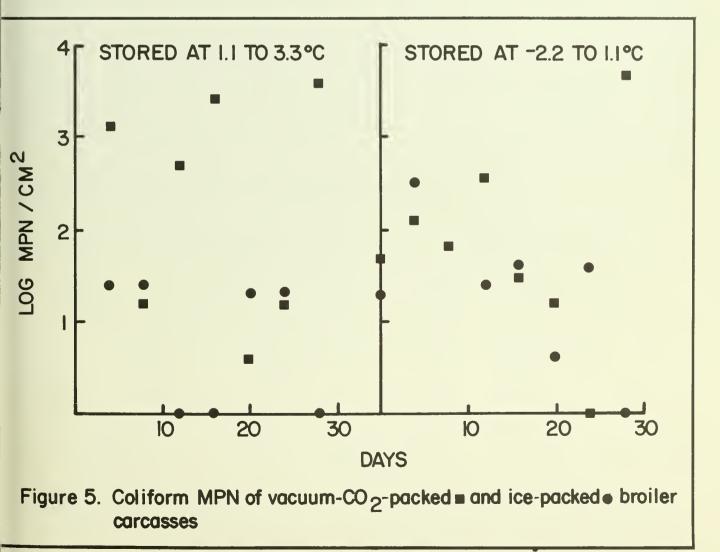
The effects of storage emperature on the growth of nicroorganisms on gizzards liffered from the effects on broiler carcasses. The stationary phase nicrobial counts for the gizzards stored at -2.2 to 1.1°C were lower han those stored at 1.1 to 3.3°C, and growth of microorganisms in ce-packed gizzard samples stored at -2.2 to 1.1°C was faster than on gizzards stored at 1.1 to 3.3°C.

In most cases, the coliform IPN's of vacuum- CO_2 – packaged amples remained low and were lightly higher than those of the cepacked controls (Figure 5). The ata supported the findings of Vabeck *et al* (1968) and Arafa and Chen (1976) but contradict those of

Sander and Soo (1978), who indicated that coliforms grew rapidly on ice-packed chickens and that the presence of CO $_2$ significantly reduced the rate of growth.

Percentages of drip in vacuum-CO₂ – packaged broiler carcasses increased slightly as storage time increased (Figure 6). Storage temperature affected the drip formation of packaged broiler carcasses. Vacuum-CO₂ – packaged carcasses stored at 1.1 to $3,3^{\circ}$ C generally had similar or slightly higher drip percentages than those stored at-2.2 to 1.1° C. The increase in drip formation of vacuum-CO₂– packaged broiler carcasses was found to be greater than that reported by Lentz and Rooke (1958) and Baker (1959). These researchers indicated that the greatest loss of moisture under refrigerated storage of packaged broiler carcasses occurred within the first 2 hrs and stabilized ater 24 hrs. A slight reabsorption of drip was observed after 24 days of refrigerated storage at both temperatures. This reabsorption of drip agreed with Arafa and Chen (1978).

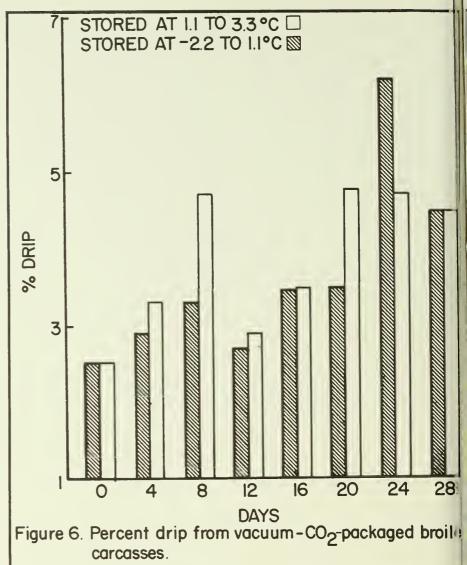
The incidence of major microbial species when broiler carcasses arrived in the laboratory was similar for the samples packed by both methods. Gram positive cocci were the dominant microorganism followed by members of *Acinetobacter* and oxidative



Moraxella (Table 1). The vacuum-CO₂ – packaged broiler carcasses had higher *Acinetobacter* counts than the ice-packed controls. In addition, no Gram positive rod type microorganisms, *Micrococcus, Pseudomonas* or fermentative *Flavobacterium* were isolated from the vacuum-CO₂– packed samples. Higher percentages of gram positive cocci on fresh broiler carcasses also were reported by Arafa and Chen (1977).

The initial incidence of the major microbial group was similar for the ice-packed and the vacuum-CO₂packaged broilers, but major groups of microflora responsible for spoilage of these two types of broiler samples were different (Table 2). The major microbial groups isolated from the spoiled ice packaged samples were members of the Pseudomonas genus, but Lactobacillus species were found to be the dominant microflora on the vacuum-CO₂ - packaged samples after 28 days of storage at 1.1 to 3.3°C.

Species of *Pseudomonas* also were isolated from the vacuum- CO_2 – packaged samples after 28 days of storage at 1.1 to 3.3°C, but



Gram-stain	Ice packed		Vacuum-CO ₂ packed		
and Morphology	No. of isolates	% of isolates	No. of isolates	% of isolates	Microbial groups
G+ cocci	34	57.6	29	52.7	Staphylococcus
	31	52.5	29	52.7	Micrococcus
	3	5.0			
G+ rod	4	6.7			
	2	3.3			B (Lactobacillus s
	2	3.3			Others
G- rod	21	35.5	26	47.2	
	2	3.3			E (Pseudomonas sp.
	2	3.3	2	3.6	J (Moraxella)
	9	15.2	18	32.7	K (Acinetobacter)
	5	8.4	4	7.2	L (<u>Moraxella</u> - oxidative)
	1	1.6			M (<u>Flavobacterium</u> - fermentative)
	2	3.3	2	3.6	N (Flavobacterium)

Gram-stain and Morphology	Ice packed		Vacuum-CO ₂ packed		
	No. of isolates	% of isolates	No. of isolates	% of isolates	Microbial groups
G+ rod	2	3.2	40	74.0	
	1	1.6	40	74.0	B (Lactobacillus sp.)
	1	1.6	-	-	Others
G- rod	59	96.7	14	25.9	
	57	93.4	12	22.2	E (Pseudomonas sp.)
	2	3.3	-	-	L (Moraxella-
	-	-	2	3.7	P (Putrefaciens)

Incidence of spoilage microflora on ice-packed and vacuum-CO₂-packed, spoiled Table 2. broilers1,

the percent Pseudomonas was much lower than for the ice-packaged samples (22.2% vs 93.4%). No oxidative Moraxella were isolated from the vacuum-CO₂ - packaged samples. The high incidence of Pseudo- days of storage at 2-4° were monas species on the spoiled ice-

incidence of with results reported by Wabeck et al (1968) and Arafa and Chen (1975). Arafa and Chen (1975) reported that 95.5% of the microorganisms in vacuumpackaged broiler parts after 28 members of *Enterobacter* species packed broiler carcasses agreed while the other 4.1% were members

of Pseudomonas. Results of this study indicated that the presence of CO_2 resulted in the growth of Lactobacillus species instead of Enterobactor species in the vacuum-CO₂ - packaged broiler carcasses.

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